

### **COMMENTARY**

## EB 1089, a Novel Vitamin D Analog with Strong Antiproliferative and Differentiation-Inducing Effects on Target Cells

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**ABSTRACT.** The physiologically active form of vitamin D,  $1\alpha$ ,25-dihydroxyvitamin D<sub>3</sub>, plays an important role not only in the establishment and maintenance of calcium metabolism, but also in regulating cell growth and differentiation. As the clinical usefulness of  $1\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> is limited by its tendency to cause hypercalcemia, new analogs with a better therapeutic profile have been synthesized. One of these new synthetic vitamin D analogs is EB 1089, which is characterized by an altered side chain structure featuring 26,27-dimethyl groups and two double bonds. This analog has been shown to be more potent than 1,25-dihydroxyvitamin D<sub>3</sub> in inhibiting proliferation, stimulating differentiation, and inducing apoptosis in a number of different cell types, including cancer cells. Despite being more potent than  $1\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> with respect to its cell regulatory effects, EB 1089 displays weaker calcemic side-effects. These characteristics make EB 1089 a potentially useful compound for the treatment of a diversity of clinical disorders, including cancer and metabolic bone diseases. A promising phase I study with EB 1089 in patients with advanced breast and colon cancer has already been carried out, and more clinical trials evaluating the clinical effectiveness of EB 1089 in other types of cancer are in progress.

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The field of vitamin D research has gained much attention due to the fact that this hormone is involved in essential cell regulatory processes such as proliferation, differentiation, and apoptosis in a number of different cell types, including cancer cells. Vitamin D has long been known to play a key role in the establishment and maintenance of calcium homeostasis. Besides this classical effect on calcium metabolism,  $1\alpha,25$ -dihydroxyvitamin  $D_3$  ( $1\alpha,25$ (OH)<sub>2</sub> $D_3$ , calcitriol) has also been found to exert cell regulatory effects that are mediated via binding to specific receptors in the target cells. During the last 15 years, numerous reports have emerged that show the presence of VDR§ in almost every cell type in the human body. Epithelial tissues such as liver, kidney, thyroid, adrenal, gastrointestinal tract, and breast cells have all been shown to possess VDR. Moreover, pituitary, parathyroid, pancreas, bone, muscle, and skin cells, as well as a number of different cancer cell types and

some activated cells of the immune system, also express high affinity receptors for  $1\alpha,25$ -dihydroxyvitamin  $D_3$  [1–3]. VDR abundance appears to be regulated by growth factors and activators of the major signalling systems in association with changes in cell proliferation and differentiation. Among others, estrogens, glucocorticoids, retinoic acid, parathyroid hormone, and  $1\alpha,25$ -dihydroxyvitamin  $D_3$  itself have been found to be capable of regulating the levels of VDR or mRNA of VDR [4–7]. At present, it is well documented that the biological function of  $1\alpha,25$ -dihydroxyvitamin  $D_3$ , in cells that are not directly associated with calcium metabolism, is to regulate cell growth and differentiation by its ability to inhibit proliferation, stimulate differentiation, and induce apoptosis [1, 2].

The exact mechanism of action of  $1\alpha$ ,25-dihydroxyvitamin  $D_3$  is not fully elucidated, but it is well accepted that the main effects of the compound are mediated by its binding to specific high affinity intracellular receptors that belong to the superfamily of steroid receptors. The receptorligand complex functions as a transcription factor by interacting with specific DNA sequences (response elements) in the promoter regions of the primary responding genes, leading to either activation or suppression of gene transcription [1, 8–10]. To date, several natural VDREs have been identified, in which the core binding motifs of six nucleotides appear to be arranged into either direct repeats or inverted palindromes with a number of spacing

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<sup>§</sup> Abbreviations: AUC∞, area under serum level/time curve; DBP, vitamin D binding protein; EB 1089, 1α,25-dihydroxy-22,24-diene-24,26,27-trihomovitamin D₃; EGF, epidermal growth factor; IGF-1, insulin-like growth factor-1; RAR, retinoic acid receptor; RXR, retinoid X receptor; TGF-β, transforming growth factor-β; TR, thyroid hormone receptor; VDR, vitamin D receptor(s); and VDRE, vitamin D response element.

nucleotides. Recent evidence suggests that the VDR binds to these response elements either as a homodimer or as a heterodimer with other nuclear receptors, preferentially with the RXR, but also with the RAR, and the TR [8]. In this way, multiple combinations and a number of different signalling pathways are possible, which may explain the great diversity of vitamin D actions.

Apart from this genomic pathway, a number of investigations have indicated the presence of another, nongenomic, pathway, in which  $1\alpha,25$ -dihydroxyvitamin  $D_3$  binds to a membrane-associated receptor that has ligand binding properties different from those of the classical vitamin D receptor described above. This receptor-ligand complex is thought to act via regulation of voltage-sensitive calcium channels, which then initiate various biological responses [3].

The broad distribution of VDR in the human body and the fact that  $1\alpha$ ,25-dihydroxyvitamin  $D_3$  is able to affect cell growth and differentiation make this hormone a potentially useful agent in the treatment of diseases such as cancer, psoriasis, and immunological disorders [2]. However, the major drawback of using  $1\alpha$ ,25-dihydroxyvitamin  $D_3$  as a therapeutic agent is that it causes hypercalcemia. Therefore, much effort has been put into the search for new vitamin D analogs with potent cell regulatory effects, but with weaker calcemic side-effects than  $1\alpha$ ,25-dihydroxyvitamin  $D_3$ .

#### **BASIC ASPECTS OF EB 1089**

One of the new promising analogs is EB 1089, which has been shown to be approximately 50-200 times more potent than  $1\alpha,25$ -dihydroxyvitamin  $D_3$  with respect to regulation of cell growth and differentiation in almost every cell type studied [11–16]. Compared with  $1\alpha,25$ -dihydroxyvitamin  $D_3$ , EB 1089 has an altered side chain structure that is characterized by an extra carbon atom (24a) and two double bonds at positions C22 and C24, respectively, and 26, 27 dimethyl groups (Fig. 1). It has been suggested that the introduction of double bonds in the side chain inhibits the hydroxylation of C23 or C24a, leading to a metabolically more stable compound [13, 17]. This hypothesis has been further supported by recent pharmacokinetic investigations that have demonstrated that EB 1089, in contrast to

FIG. 1. Structure of  $1\alpha,25$ -dihydroxyvitamin  $D_3$   $(1\alpha,25(OH)_2D_3)$  and the side-chain structure of EB 1089.

1α,25(OH)<sub>2</sub>D<sub>3</sub>

(6-s-trans conformation)

most other synthetic vitamin D analogs, is metabolically more stable, having a serum half-life comparable to that of  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> (Table 1). On the other hand, the AUC∞, which indicates the concentration and persistence of a compound in the serum, has been found to be considerably higher for  $1\alpha,25$ -dihydroxyvitamin  $D_3$  than for EB 1089 (Table 1). This is apparently due to the lower affinity of EB 1089 for the DBP compared with  $1\alpha,25$ dihydroxyvitamin D<sub>3</sub>, as the DBP binding affinity of vitamin D analogs correlates with their serum levels or the volume of distribution, which then influences the clearance of the compounds [13, 18]. Moreover, the effect of EB 1089 on calcium metabolism in vivo in rats has been shown to be approximately 50% lower than that of 1a,25-dihydroxyvitamin D<sub>3</sub> [13, 19]. This may also be due, in part, to its relatively low binding affinity for DBP and the increased clearance, but other mechanisms such as tissue-specific differences in the binding affinities of 1a,25-dihydroxyvitamin D<sub>3</sub> and EB 1089 for the VDR, and different signalling pathways have also been suggested to contribute to the reduced calcemic activity of EB 1089 [11, 17, 20, 21].

The majority of synthetic vitamin D analogs appear to bind with a lower affinity to VDR than  $1\alpha,25$ -dihydroxyvi-

TABLE 1. Pharmacological data on 10,25-dihydroxyvitamin D<sub>3</sub> (10,25(OH)<sub>2</sub>D<sub>3</sub>) and EB 1089

Compound	Binding to DBP 50% displacement* (M)	Serum half-life in vivo (T <sub>1/2</sub> )* (hr)	AUC∞* (ng/mL·hr)	Calcemic activity in vivo† (%)
EB 1089	$7.9 \times 10^{-6}$ $1.5-6.0 \times 10^{-7}$	2.1	255	50
1α,25(OH) <sub>2</sub> D <sub>3</sub>		2.2	7355	100

The binding affinity for DBP was assessed by displacing  $[^3H]$ -1,25(OH)<sub>2</sub>D<sub>3</sub> from a DBP-Affi-Gel.  $T_{1/2}$  and AUC $\infty$  were measured following a single i.v. dose of 200  $\mu$ g/kg to rats, and the calcemic activity was determined after p.o. administration for 7 days in rats.

<sup>\*</sup> Data were obtained from Ref. 18.

<sup>†</sup> Data were obtained from Ref. 13.

TABLE 2. Antiproliferative effects of $1\alpha,25$ -dihydroxyvitamin $D_3$ ( $1\alpha,25$ (OH) $_2D_3$ ) and EB 1089 on three different human cell lines in vitro						
	vmnhoma cells*	MCF-7 breast cancer cells†	HaCaT keratinocytes*			

Compound	U937 lymphoma cells* (IC <sub>50</sub> ) (M)	MCF-7 breast cancer cells† (IC <sub>50</sub> ) (M)	HaCaT keratinocytes* (IC <sub>50</sub> ) (M)
EB 1089 1α25(OH) <sub>2</sub> D <sub>3</sub>	$5.8 \times 10^{-10}$ $4.6 \times 10^{-8}$	$ \begin{array}{c} 2.3 \times 10^{-10} \\ 1.0 \times 10^{-8} \end{array} $	$7.9 \times 10^{-10} \\ 7.1 \times 10^{-8}$

<sup>\*</sup> Data obtained from Ref. 13.

tamin D<sub>3</sub>, as demonstrated by displacement studies using isolated VDR from different cell sources [11, 16, 22, 23]. Despite being more potent than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> in the regulation of cell growth and differentiation, EB 1089 was found to have a reduced binding affinity for VDR compared with  $1\alpha,25$ -dihydroxyvitamin  $D_3$  in most of the cell systems studied [11, 13, 16, 19, 24]. Recent evidence suggests, however, that the biological activity of vitamin D compounds is not determined by the receptor binding affinity alone. Other important factors such as the mode of interaction of the compounds with the VDR, their ability to form receptor-dimer complex, and their preference for specific VDRE types all contribute to the activation of VDR-mediated transcription of the effector genes, and thus determine the biological activity [8, 11, 14, 20, 25]. Interestingly, it has been demonstrated that EB 1089 is clearly more efficient than  $1\alpha,25$ -dihydroxyvitamin  $D_3$  in stimulating gene activity, presumably due to its ability to enhance the stability of the occupied receptor, to stimulate the formation of the dimerization of VDR with RXR, and/or to cause a stronger, longer-lasting binding of the receptor complex to VDRE [8, 11, 14, 20, 25]. Moreover, recent investigations have suggested that EB 1089, in contrast to  $1\alpha,25$ -dihydroxyvitamin  $D_3$ , preferentially activates a VDRE (IP9-type), which seems to be more closely related to the regulation of cell growth than to the classical actions of vitamin D, i.e. the effects on calcium metabolism [15, 26].

# EFFECTS OF EB 1089 ON EARLY GENOMIC CHANGES

Today more than 50 genes have been reported to be sensitive to vitamin D, including several genes that are known to be involved in the regulation of cell proliferation and differentiation [1, 8]. Very recently it has been shown that one of the key regulators in the cell cycle, the cyclin-dependent kinase-inhibitor p21 WAFI,CIPI, contains a VDRE and is transcriptionally induced by  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> [27, 28]. Further studies at the protein level have demonstrated that EB 1089 is more potent than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> in up-regulating the p21 protein, suggesting that EB 1089 acts as a more effective blocker of cell transition from G<sub>1</sub> to S-phase in the cell cycle than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> [29]. Another newly identified primary vitamin D responding gene of particular

importance for cell cycle progression is the gene encoding for c-fos [30]. Previous investigations have shown that this gene is transcriptionally regulated by vitamin D, and that EB 1089 is approximately 50 times more potent in terms of up-regulating the expression of c-fos mRNA in MCF-7 breast cancer cells, compared with  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> [19]. Moreover, EB 1089 has been found to exert stronger effects than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> on several early responding genes. These include the proto-oncogene c-myc, the growth regulator gene TGF- $\beta$ , and the apoptosis related gene TRPM-2 (clustrin), which all show altered mRNA expression in breast cancer cells treated with EB 1089 [19, 29, 31].

#### EFFECTS OF EB 1089 ON NEOPLASTIC CELLS

It is well documented that  $1\alpha,25$ -dihydroxyvitamin  $D_3$ inhibits cell growth and promotes cell differentiation in several different cancer cell types in vitro. These include cells derived from tumors of the breast, prostate, pancreas, colon, bladder, thyroid, pituitary, and skin as well as melanoma, glioma, neuroblastoma, leukemia, and lymphoma cells [1, 22, 23, 32-34]. In addition to its antiproliferative and differentiation-inducing effects, 1α,25-dihydroxyvitamin D<sub>3</sub> has been found to induce apoptosis and to decrease the invasiveness of a number of different cancer cells in vitro [29, 32, 33, 35, 36\*]. In vivo, inhibition of the development of metastases, regression of tumors, inhibition of angiogenesis, prevention of malignancy-associated hypercalcemia, and prolongation of survival time have been observed in tumor-bearing animals treated with the hormone [29, 36-39].

#### In Vitro Effects

In most cancer cell systems in vitro, EB 1089 has been found to be approximately 50-200 times more potent than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> with respect to inhibition of cell proliferation [11, 13–16, 19, 33, 40] (Table 2). This may be due, in part, to its more pronounced effects on cell cycle regulators like c-myc, c-fos, p21, and p53, but also

<sup>†</sup> Data obtained from Ref. 19.

<sup>\*</sup> Hoelting T, Duh QY, Clark OH and Herfarth C, Vitamin D and its analog EB-1089 inhibit growth, invasion and protease activity in follicular thyroid cancer cells *in vitro*. 10th International Congress on Endocrinology, San Francisco, CA, 12–15 June, 1996, Abstr. 160.

modulation of the levels of estrogen receptors and of TGF-β type II receptors seems to contribute to the antiproliferative effect of the compound. In MCF-7 breast cancer cells, EB 1089 has been shown to cause a more potent reduction of the estrogen receptor concentration than  $1\alpha,25$ -dihydroxyvitamin  $D_3$ , leading to a more effective block of the mitogenic effect of estradiol [41-43]. In addition, besides being able to increase the level of TGF-β [31], a recent study has indicated that TGF-B type II receptors are up-regulated in MCF-7 cells in response to treatment of the cells with EB 1089, resulting in an increase in TGF-β autocrine negative growth activity [44]. Also, the mitogenic activity of insulin, IGF-1, and EGF in human breast cancer cells has been shown to be blocked by vitamin D analogs, with EB 1089 being more potent than  $1\alpha,25$ dihydroxyvitamin D<sub>3</sub> [40, 45].

Another important factor for the regulation of cell growth is apoptosis, and a number of recent studies have indicated that EB 1089 is more efficient than  $1\alpha,25$ dihydroxyvitamin D<sub>3</sub> in inducing this process. In MCF-7 human breast cancer cells, EB 1089 has proven to be approximately 50-100 times more potent than  $1\alpha,25$ dihydroxyvitamin D<sub>3</sub> in stimulating the expression of TRPM-2 and cathepsin B and in down-regulating the expression of bcl-2, which all serve as early markers of apoptosis [29, 43, 46]. Moreover, an increased fragmentation of genomic DNA was seen when the cells were treated with EB 1089, suggesting that active cell death is occurring in the cells [29, 46]. Interestingly, the apoptosis-inducing effect of EB 1089 could be further augmented by cotreatment of the cells with EB 1089 and 9-cis-retinoic acid [46]. Co-operative effects of the combination of EB 1089 with retinoids have also been observed in other cell types, including human pancreatic cells, human breast cancer cells, and in certain colon carcinoma cell lines, where the antiproliferative effect of EB 1089 was shown to be enhanced in the presence of 9-cis- or all-trans-retinoic acid [46-48]. In leukemia cells, the growth-reducing effects of EB 1089 were accompanied by a marked increase in the expression of differentiation markers like CD11b, CD14, and CD18 [49]. This differentiation-inducing effect and the antiproliferative effect of EB 1089 on these cells were increased further by simultaneous treatment of the cells with EB 1089 and 9-cis- or all-trans-retinoic acid [49].

Differentiation-inducing effects in response to treatment of the cells with EB 1089 have also been documented in other cancer cell types. These include transformed human keratinocytes, human prostate cancer cells, and human neuroblastoma cells, in which up-regulation of, respectively, fillagrin and cytokeratin K10, prostate specific antigen, and acetylcholinesterase activity have been observed [33, 34, 50]. In human neuroblastoma cells, the increased activity of acetylcholinesterase was found to be accompanied by a decreased expression of N-myc, which is known to precede neuroblastoma differentiation. Moreover, the N-myc expression was also shown to be correlated directly with a reduced ability of the cells to invade an artificial

basement membrane [33]. The anti-invasive effect of EB 1089 was further confirmed by other studies using human breast cancer cells and human follicular thyroid cancer cells [33–35]. Interestingly, these studies have proven that the anti-invasive effect of vitamin D analogs is not associated directly with their growth-inhibiting potential. Other mechanisms, such as decreased expression of collagenases and a reduction of the laminin receptor expression, may also be involved [11, 34, 35, 51].

### In Vivo Effects

The studies described above have demonstrated that EB 1089 has potent cell regulatory effects in vitro, which could be of great interest for the treatment of neoplastic diseases. As its calcemic activity was found to be approximately 50% lower than that of  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> [13, 19], EB 1089 has been selected for further investigations in vivo and has been tested in a series of assays. In rats with mammary tumors induced by nitrosomethylurea, oral treatment of the animals with EB 1089 at 0.5 µg/kg/day for 4 weeks was shown to result in a significant inhibition of tumor growth and a significant regression of the tumor size without changes in serum calcium levels [11, 29]. In addition, in rats bearing the Leydig tumor H-500, EB 1089 was found to prolong the survival time and to prevent the induction of malignancy-associated hypercalcemia when administered at non-calcemic doses [11, 37].

EB 1089 is currently undergoing clinical evaluation in patients suffering from a variety of cancer types. A phase I study in patients with advanced breast and colon cancer has indicated that EB 1089 can be administered at relatively high doses without causing hypercalcemia or other toxicities [52].

#### EFFECTS OF EB 1089 ON BONE CELLS

Vitamin D has long been known to affect bone growth and mineralization and serve as an important agent for the treatment of metabolic bone diseases. In vitro, 1α,25dihydroxyvitamin D<sub>3</sub> has proven to be a potent regulator of bone cell functions, and it is capable of inducing differentiation of the two major bone cells, osteoblasts and osteoclasts [53]. VDRs have been demonstrated in the osteoblasts, while osteoclasts, which are derived from immature bone marrow cells, do not contain VDR [9, 53]. The differentiation-inducing effect of 1α,25-dihydroxyvitamin D<sub>3</sub> in osteoclasts is therefore indirect and is apparently mediated by a factor that is released from the osteoblastic cells in response to treatment of the cells with  $1\alpha,25$ dihydroxyvitamin D<sub>3</sub> [9, 53]. Differentiation of the osteoclast precursors results in an increased bone resorbing activity of the cells by affecting the matrix attachment phase of the resorption process [54]. In the osteoblasts,  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> has been shown to regulate the expression of several maturation associated proteins such as alkaline phosphatase, collagen, osteocalcin, osteopontin,

matrix Gla-protein, and the third component of complement (C3) [53]. Some of these effects are mediated through changes in the local production of cytokines such as TGF- $\beta$ , interleukin-4, and EGF, and their receptors [55–57].

EB 1089 and  $1\alpha$ ,25-dihydroxyvitamin  $D_3$  were found to be equipotent stimulators of osteoclast recruitment in murine marrow cells in vitro [24]. However, in in vitro resorption experiments using bone cells from fetal mice, EB 1089 appeared to be approximately 10 times more potent than  $1\alpha$ ,25-dihydroxyvitamin  $D_3$  in stimulating the bone resorption activity of the cells. This stimulatory effect could be reduced considerably by addition of the antiestrogen, tamoxifen, suggesting a role for such a combination therapy in reducing the risk of skeletal metastases in cancer patients treated with vitamin D analogs [58].

Other *in vitro* studies using MG-63 osteosarcoma cells have shown that EB 1089 inhibits cell proliferation more potently than  $1\alpha,25$ -dihydroxyvitamin  $D_3$  and that it causes a marked increase in the synthesis of procollagen type I and osteocalcin [14, 16]. Moreover, EB 1089 appears to be more potent than  $1\alpha,25$ -dihydroxyvitamin  $D_3$  with respect to the stimulation of alkaline phosphatase activity and to increasing the level of VDR [14, 16]. One explanation for the stronger effects on bone cells of EB 1089 than  $1\alpha,25$ -dihydroxyvitamin  $D_3$  may be the fact that the binding of VDR to the composite AP-1 plus VDRE promoter region of the human osteocalcin gene after EB 1089 is stronger and longer lasting than after  $1\alpha,25$ -dihydroxyvitamin  $D_3$  treatment [14, 16].

# SUMMARY AND DIRECTIONS FOR FUTURE RESEARCH

On the basis of both in vitro and in vivo studies, EB 1089 emerges as one of the promising new vitamin D analogs. It has a conjugated double-bond side chain structure, which probably makes it metabolically more stable than the parent compound,  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub>. EB 1089 belongs to the group of vitamin D analogs characterized by having a low affinity for the DBP, a relatively long serum half-life, and weak calcemic side-effects compared with those of  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub>. It is more potent than  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub> with respect to regulation of cell growth and differentiation, probably due to the fact that its effects on target cell DNA, mediated by the vitamin D receptor, are stronger and longer lasting than those of  $1\alpha,25$ -dihydroxyvitamin D<sub>3</sub>. Moreover, EB 1089 has also been found to exert potent effects on bone metabolism, affecting both osteoblastic and osteoclastic cells.

In basic research, it is anticipated that the X-ray structures of VDR and DBP will be determined in the near future. Knowledge of these structures may help clarify their structure-function relationships and thus greatly assist in the development and search for new vitamin D analogs with specific pharmacological profiles. Thus, in the future, the use of vitamin D analogs may hopefully extend beyond its classical role in bone diseases, to encompass new areas in

the field of neoplastic diseases either as a monotherapy or in combination with other anticancer agents.

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#### References

- Hannah SS and Norman AW, 1α,25(OH)<sub>2</sub> Vitamin D<sub>3</sub>regulated expression of the eukaryotic genome. Nutr Rev 52:
  376–382, 1994.
- 2. Holick MF, Noncalcemic actions of 1,25-dihydroxyvitamin D<sub>3</sub> and clinical applications. *Bone* 17: 107S–111S, 1995.
- Norman AW, Bishop JE, Collins ED, Seo E-G, Satchell DP, Dormanen MC, Zanello SB, Farach-Carson MC, Bouillon R and Okamura WH, Differing shapes of 1α,25-dihydroxyvitamin D<sub>3</sub> function as ligands for the D-binding protein, nuclear receptor and membrane receptor: A status report. J Steroid Biochem Mol Biol 56: 13–22, 1996.
- Mahonen A and Mäenpää PH, Steroid hormone modulation of vitamin D receptor levels in human MG-63 osteosarcoma cells. Biochem Biophys Res Commun 205: 1179–1186, 1994.
- 5. Darwish H and DeLuca HF, Vitamin D regulated gene expression. Crit Rev Eukaryot Gene Expr 3: 89-116, 1993.
- Duncan WE, Glass AR and Wray HL, Estrogen regulation of the nuclear 1,25-dihydroxyvitamin D<sub>3</sub> receptor in rat liver and kidney. *Endocrinology* 129: 2318–2324, 1991.
- Chen TL and Feldman D, Retinoic acid modulation of 1,25(OH)<sub>2</sub> vitamin D<sub>3</sub> receptors and bioresponse in bone cells: Species difference between rat and mouse. *Biochem Biophys Res Commun* 132: 74–80, 1985.
- Carlberg C, Mechanisms of nuclear signalling by vitamin D<sub>3</sub>. Interplay with retinoid and thyroid hormone signalling. Eur J Biochem 231: 517–527, 1995.
- Christakos S, Raval-Pandya M, Wernyj PP and Yang W, Genomic mechanisms involved in the pleiotropic actions of 1,25-dihydroxyvitamin D<sub>3</sub>. Biochem J 316: 361–371, 1996.
- Peleg S, Sastry M, Collins ED, Bishop JE and Norman AW, Distinct conformational changes induced by 20-epi analogues of 1α,25-dihydroxyvitamin D<sub>3</sub> are associated with enhanced activation of the vitamin D receptor. J Biol Chem 270: 10551–10558, 1995.
- 11. Binderup L, Carlberg C, Kissmeyer A-M, Latini S, Mathiasen IS and Mørk Hansen C, The need for new vitamin D analogues: Mechanisms of action and clinical applications. In: Vitamin D—A Pluripotent Steroid Hormone: Structural Studies, Molecular Endocrinology and Clinical Applications (Eds. Bouillon R, Norman AW and Thomasset M), pp. 55–63. Walter de Gruyter, Berlin, 1994.
- 12. Colston KW, Vitamin D and breast cancer: The therapeutic potential of new vitamin D analogues. *Endocr Relat Cancer* 2: 187–201, 1995.
- Kissmeyer A-M, Binderup E, Binderup L, Mørk Hansen C, Rastrup Andersen N, Makin HLJ, Schroeder NJ, Shankar VN and Jones G, Metabolism of the vitamin D analog EB 1089: Identification of in vivo and in vitro liver metabolites and their biological activities. Biochem Pharmacol 53: 1087–1097, 1997.
- 14. Mahonen A, Jääskeläinen T and Mäenpää PH, A novel vitamin D analog with two double bonds in its side chain. A potent inducer of osteoblastic cell differentiation. *Biochem Pharmacol* **51:** 887–892, 1996.
- Nayeri S, Danielsson C, Kahlen J-P, Schräder M, Mathiasen IS, Binderup L and Carlberg C, The anti-proliferative effect of vitamin D<sub>3</sub> analogues is not mediated by inhibition of the

- AP-1 pathway, but may be related to promoter selectivity. Oncogene 11: 1853–1858, 1995.
- van den Bemd GJCM, Pols HAP, Birkenhäger JC, Kleinekoort WMC and van Leeuwen JPTM, Differential effects of 1,25-dihydroxyvitamin D<sub>3</sub>-analogs on osteoblast-like cells and on in vitro bone resorption. J Steroid Biochem Mol Biol 55: 337–346, 1995.
- Roy S, Martel J and Tenenhouse HS, Comparative effects of 1,25-dihydroxyvitamin D<sub>3</sub> and EB 1089 on mouse renal and intestinal 25-hydroxyvitamin D<sub>3</sub>-24-hydroxylase. J Bone Miner Res 10: 1951–1959, 1995.
- 18. Kissmeyer A-M, Mathiasen IS, Latini S and Binderup L, Pharmacokinetic studies of vitamin D analogues: Relationship to vitamin D binding protein (DBP). *Endocrine* 3: 263–266, 1995.
- 19. Mathiasen IS, Colston KW and Binderup L, EB 1089, a novel vitamin D analogue, has strong antiproliferative and differentiation inducing effects on cancer cells. *J Steroid Biochem Mol Biol* **46:** 365–371, 1993.
- Carlberg C, Mathiasen IS, Saurat J-H and Binderup L, The 1,25-dihydroxyvitamin D<sub>3</sub> (VD) analogues MC 903, EB 1089 and KH 1060 activate the VD receptor: Homodimers show higher ligand sensitivity than heterodimers with retinoid X receptors. J Steroid Biochem Mol Biol 51: 137–142, 1994.
- Norman AW, Sergeev IN, Bishop JE and Okamura WH, Selective biological response by target organs (intestine, kidney and bone) to 1,25-dihydroxyvitamin D<sub>3</sub> and two analogues. Cancer Res 53: 3935–3942, 1993.
- Bouillon R, Okamura WH and Norman AW, Structurefunction relationships in the vitamin D endocrine system. Endocr Rev 16: 200–257, 1995.
- 23. Mørk Hansen C, Mathiasen IS and Binderup L, The anti-proliferative and differentiation-inducing effects of vitamin D analogues are not determined by the binding affinity for the vitamin D receptor alone. J Invest Dermatol Symp Proc 1: 44–48, 1996.
- Wiberg K, Ljunghall S, Binderup L and Ljunggren Ö, Studies on two new vitamin D analogs, EB 1089 and KH 1060: Effects on bone resorption and osteoclast recruitment *in vitro*. *Bone* 17: 391–395, 1995.
- 25. Mathiasen IS, MacKay AG, Colston KW and Binderup L, EB 1089, a vitamin D analogue, induces the vitamin D receptor to form selective receptor complexes depending on cell type. In: Vitamin D—A Pluripotent Steroid Hormone: Structural Studies, Molecular Endocrinology and Clinical Applications (Eds. Bouillon R, Norman AW and Thomasset M), pp. 255–256. Walter de Gruyter, Berlin, 1994.
- Mørk Hansen C, Danielsson C and Carlberg C, The potent anti-proliferative effect of 20-epi analogues of 1,25-dihydroxyvitamin D<sub>3</sub> in human breast-cancer MCF-7 cells is related to promoter selectivity. *Int J Cancer* 67: 739–742, 1996
- 27. Jiang H, Lin J, Su Z-Z, Collart FR, Huberman E and Fisher PB, Induction of differentiation in human promyelocytic HL-60 leukemia cells activates p21, WAF1/CIP1, expression in the absence of p53. Oncogene 9: 3397–3406, 1994.
- Liu M, Lee M-H, Cohen M, Bommakanti M and Freedman LP, Transcriptional activation of the cdk inhibitor p21 by vitamin D<sub>3</sub> leads to the induced differentiation of the myelomonocytic cell line U937. Genes Dev 10: 142–153, 1996.
- James SY, MacKay AG and Colston KW, Effects of 1,25dihydroxyvitamin D<sub>3</sub> and its analogues on induction of apoptosis in breast cancer cells. J Steroid Biochem Mol Biol 58: 395–401, 1996.
- 30. Candeliere GA, Jurutka PW, Haussler MR and St-Arnaud R, A composite element binding the vitamin D receptor, retinoid X receptor α, and a member of the CTF/NF-1 family of

- transcription factors mediates the vitamin D responsiveness of the c-fos promoter. Mol Cell Biol 16: 584-592, 1996.
- Koli K and Keski-Oja J, 1,25-Dihydroxyvitamin D<sub>3</sub> enhances the expression of transforming growth factor β1 and its latent form binding protein in cultured breast carcinoma cells. Cancer Res 55: 1540–1546, 1995.
- 32. Baudet C, Chevalier G, Naveilhan P, Binderup L, Brachet P and Wion D, Cytotoxic effects of 1α,25-dihydroxyvitamin D<sub>3</sub> and synthetic vitamin D<sub>3</sub> analogues on a glioma cell line. *Cancer Lett* 100: 3–10, 1996.
- Moore TB, Koeffler HP, Yamashiro JM and Wada RK, Vitamin D<sub>3</sub> analogs inhibit growth and induce differentiation in LA-N-5 human neuroblastoma cells. Clin Exp Metastasis 14: 239–245, 1996.
- Skowronski RJ, Peehl DM and Feldman D, Actions of vitamin D<sub>3</sub> analogs on human prostate cancer cell lines: Comparison with 1,25-dihydroxyvitamin D<sub>3</sub>. Endocrinology 136: 20–26, 1995.
- Mørk Hansen C, Frandsen TL, Brünner N and Binderup L, 1α,25-Dihydroxyvitamin D<sub>3</sub> inhibits the invasive potential of human breast cancer cells in vitro. Clin Exp Metastasis 12: 195–202, 1994.
- 36. Young MRI, Halpin J, Hussain R, Lozano Y, Djordjevic A, Devata S, Matthews JP and Wright MA, Inhibition of tumor production of granulocyte-macrophage colony-stimulating factor by 1α,25-dihydroxyvitamin D<sub>3</sub> reduces tumor motility and metastasis. *Invasion Metastasis* 13: 169–177, 1993.
- 37. Haq M, Kremer R, Goltzman D and Rabbani SA, A vitamin D analogue (EB1089) inhibits parathyroid hormone-related peptide production and prevents the development of malignancy-associated hypercalcemia in vivo. J Clin Invest 91: 2416–2422, 1993.
- 38. Oikawa T, Yoshida Y, Shimamura M, Ashino-Fuse H, Iwaguchi T and Tominaga T, Antitumor effect of 22-oxa-1α,25-dihydroxyvitamin D<sub>3</sub>, a potent angiogenesis inhibitor, on rat mammary tumors induced by 7,12-dimethylbenz(a)anthracene. *Anticancer Drugs* 2: 475–480, 1991.
- 39. Tanaka Y, Wu AYS, Ikekawa N, Iseki K, Kawai M and Kobayashi Y, Inhibition of HT-29 human colon cancer growth under the renal capsule of severe combined immunodeficient mice by an analogue of 1,25-dihydroxyvitamin D<sub>3</sub>, DD-003. Cancer Res **54**: 5148–5153, 1994.
- 40. Vink-van Wijngaarden T, Pols HAP, Buurman CJ, Birkenhäger JC and van Leeuwen JPTM, Inhibition of insulin-like growth factor-1-stimulated growth of human breast cancer cells by 1,25-dihydroxyvitamin D<sub>3</sub> and the vitamin D<sub>3</sub> analogue EB 1089. Eur J Cancer 32A: 842–848, 1996.
- 41. James SY, Mackay AG, Binderup L and Colston KW, Effects of a new synthetic vitamin D analogue, EB 1089, on the oestrogen-responsive growth of human breast cancer cells. *J Endocrinol* 141: 555–563, 1994.
- 42. Simboli-Campbell M and Welsh J, Comparative effects of 1,25(OH)<sub>2</sub>D<sub>3</sub> and EB 1089 on cell cycle kinetics in MCF-7 cells. In: Vitamin D—A Pluripotent Steroid Hormone: Structural Studies, Molecular Endocrinology and Clinical Applications (Eds. Bouillon R, Norman AW and Thomasset M), pp. 506–507. Walter de Gruyter, Berlin, 1994.
- 43. Welsh J, Induction of apoptosis in breast cancer cells in response to vitamin D and antiestrogens. *Biochem Cell Biol* 72: 537–545, 1994.
- 44. Wu GF, Sun LZ and Brattain MG, Effects of 1,25(OH)<sub>2</sub> vitamin D<sub>3</sub> and its analogue EB 1089 on regulation of TGFβ type II receptor in human breast cancer MCF-7 cells. Proc Am Assoc Cancer Res 37: 226, 1996.
- 45. Saez S, Meggouh F, Lefebre M-F, Descotes F, Pamphile R, Adam L and Crepin M, Potential direct and indirect influence of 1,25 (OH)<sub>2</sub>D<sub>3</sub> on the growth of human colonic and breast cancer. In: Vitamin D—A Pluripotent Steroid Hormone:

- Structural Studies, Molecular Endocrinology and Clinical Applications (Eds. Bouillon R, Norman AW and Thomasset M), pp. 469–473. Walter de Gruyter, Berlin, 1994.
- James SY, Mackay AG and Colston KW, Vitamin D derivatives in combination with 9-cis retinoic acid promote active cell death in breast cancer cells. J Mol Endocrinol 14: 391–394, 1995.
- 47. Kane KF, Langman MJS and Williams GR, 9-cis Retinoic acid differentially modifies vitamin D<sub>3</sub> action in two human colorectal cancer cell lines. Gut 36 (Suppl 1): A7, 1995.
- 48. Zugmaier G, Jäger R, Grage B, Gottardis MM, Havemann K and Knabbe C, Growth-inhibitory effects of vitamin D analogues and retinoids on human pancreatic cancer cells. Br J Cancer 73: 1341–1346, 1996.
- 49. James SY, Kelsey SM, Williams MA, Newland AC and Colston KW, Interactions between vitamin D derivatives, retinoids and granulocyte macrophage-colony stimulating factor in leukemic cell differentiation. In: Vitamin D—A Pluripotent Steroid Hormone: Structural Studies, Molecular Endocrinology and Clinical Applications (Eds. Bouillon R, Norman AW and Thomasset M), pp. 461–462. Walter de Gruyter, Berlin, 1994.
- Mils V, Basset-Séguin N, Molès J-P and Guilhou J-J, 1,25-Dihydroxyvitamin D<sub>3</sub> and its synthetic derivatives MC 903 and EB 1089 induce a partial tumor phenotype reversal in a skin-equivalent system. J Invest Dermatol Symp Proc 1: 87–93, 1996.
- 51. Mørk Hansen C, Arensbak B, Latini S and Binderup L, Down regulation of laminin-binding integrins by 1α,25-dihy-

- droxyvitamin D<sub>3</sub> in human melanoma cells in vitro. Cell Adhesion Commun, in press.
- Gulliford T, English J, Colston K, Sprogel P and Coombes RC, A phase 1 study of EB1089, a vitamin D analogue. Proc Am Assoc Cancer Res 37: 164, 1996.
- 53. Suda T, Takahashi N and Abe E, Role of vitamin D in bone resorption. *J Cell Biochem* **49:** 53–58, 1992.
- 54. Medhora MM, Teitelbaum S, Chappel J, Alvarez J, Mimura H, Ross FP and Hruska K, 1α,25-Dihydroxyvitamin D<sub>3</sub> up-regulates expression of the osteoclast integrin α<sub>ν</sub>β<sub>3</sub>. J Biol Chem 268: 1456–1461, 1993.
- Petkovich PM, Wrana JL, Grigoriadis AE, Heersche JNM and Sodek J, 1,25-Dihydroxyvitamin D<sub>3</sub> increases epidermal growth factor receptors and transforming growth factor-β-like activity in a bone-derived cell line. J Biol Chem 262: 13424–13428, 1987.
- 56. Finkelman RD, Linkhart TA, Mohan S, Lau K-HW, Baylink DJ and Bell NH, Vitamin D deficiency causes a selective reduction in deposition of transforming growth factor-β in rat bone: Possible mechanism for impaired osteoinduction. Proc Natl Acad Sci USA 88: 3657–3660, 1991.
- Lacey DL, Erdmann JM, Tan H-L and Ohara J, Murine osteoblast interleukin 4 receptor expression: Upregulation by 1,25-dihydroxyvitamin D<sub>3</sub>. J Cell Biochem 53: 122–134, 1993.
- 1,25-dihydroxyvitamin D<sub>3</sub>. *J Cell Biochem* **53**: 122–134, 1993.

  58. Vink-van Wijngaarden T, Birkenhäger JC, Kleinekoort WMC, van den Bemd G-JCM, Pols HAP and Leeuwen JPTM, Antiestrogens inhibit *in vitro* bone resorption stimulated by 1,25-dihydroxyvitamin D<sub>3</sub> and the vitamin D<sub>3</sub> analogs EB 1089 and KH 1090. *Endocrinology* **136**: 812–815, 1995.